



Australian Government
Department of Health

COMMUNICABLE DISEASES INTELLIGENCE

2020 Volume 44
<https://doi.org/10.33321/cdi.2020.44.4>

Australian Gonococcal Surveillance Programme Annual Report, 2018

Monica M Lahra, Rodney Enriquez and CR Robert George
for the National Neisseria Network, Australia

Communicable Diseases Intelligence

ISSN: 2209-6051 Online

This journal is indexed by Index Medicus and Medline.

Creative Commons Licence - Attribution-NonCommercial-NoDerivatives CC BY-NC-ND

© 2020 Commonwealth of Australia as represented by the Department of Health

This publication is licensed under a Creative Commons Attribution-Non-Commercial NoDerivatives 4.0 International Licence from <https://creativecommons.org/licenses/by-nc-nd/4.0/legalcode> (Licence). You must read and understand the Licence before using any material from this publication.

Restrictions

The Licence does not cover, and there is no permission given for, use of any of the following material found in this publication (if any):

- the Commonwealth Coat of Arms (by way of information, the terms under which the Coat of Arms may be used can be found at www.itsanhonour.gov.au);
- any logos (including the Department of Health's logo) and trademarks;
- any photographs and images;
- any signatures; and
- any material belonging to third parties.

Disclaimer

Opinions expressed in Communicable Diseases Intelligence are those of the authors and not necessarily those of the Australian Government Department of Health or the Communicable Diseases Network Australia. Data may be subject to revision.

Enquiries

Enquiries regarding any other use of this publication should be addressed to the Communication Branch, Department of Health, GPO Box 9848, Canberra ACT 2601, or via e-mail to: copyright@health.gov.au

Communicable Diseases Network Australia

Communicable Diseases Intelligence contributes to the work of the Communicable Diseases Network Australia.
<http://www.health.gov.au/cdna>



Communicable Diseases Intelligence (CDI) is a peer-reviewed scientific journal published by the Office of Health Protection, Department of Health. The journal aims to disseminate information on the epidemiology, surveillance, prevention and control of communicable diseases of relevance to Australia.

Editor

Cindy Toms

Deputy Editor

Simon Petrie

Design and Production

Kasra Yousefi

Editorial Advisory Board

David Durrheim,
Mark Ferson, John Kaldor,
Martyn Kirk and Linda Selvey

Website

<http://www.health.gov.au/cdi>

Contacts

Communicable Diseases Intelligence is produced by:
Health Protection Policy Branch
Office of Health Protection
Australian Government
Department of Health
GPO Box 9848, (MDP 6)
CANBERRA ACT 2601

Email:

cdi.editor@health.gov.au

Submit an Article

You are invited to submit your next communicable disease related article to the Communicable Diseases Intelligence (CDI) for consideration. More information regarding CDI can be found at:
<http://health.gov.au/cdi>.

Further enquiries should be directed to:
cdi.editor@health.gov.au.

Australian Gonococcal Surveillance Programme Annual Report, 2018

Monica M Lahra, Rodney Enriquez and CR Robert George for the National Neisseria Network, Australia

Abstract

The Australian Gonococcal Surveillance Programme (AGSP) has continuously monitored antimicrobial resistance in clinical isolates of *Neisseria gonorrhoeae* from all states and territories since 1981. In 2018, there were 9,006 clinical isolates of gonococci from public and private sector sources tested for *in vitro* antimicrobial susceptibility by standardised methods. This was the highest annual total of isolates tested since the inception of the AGSP. The current treatment recommendation for gonorrhoea, for the majority of Australia, remains dual therapy with ceftriaxone and azithromycin. Decreased susceptibility to ceftriaxone (minimum inhibitory concentration (MIC) value ≥ 0.06 mg/L) was found nationally in 1.73% of isolates. The highest proportions were reported from Tasmania and non-remote Western Australia (7.3% and 2.1% respectively). In 2018 two extensively drug-resistant isolates were reported from Queensland patients. These two isolates, with ceftriaxone MIC values of 0.50 mg/L, high-level resistance to azithromycin (MIC ≥ 256 mg/L), and resistance to penicillin and ciprofloxacin were identified and reported to the World Health Organization as isolates of international significance. Resistance to azithromycin (MIC value ≥ 1.0 mg/L) was found nationally in 6.2% of isolates, lower than the 9.3% reported in 2017, but more than double the proportion reported in 2015 (2.6%). The highest proportions were reported from the Australian Capital Territory (8.7%), Victoria (8.3%), and New South Wales (6.5%). High-level resistance to azithromycin (MIC value ≥ 256 mg/L) was reported in nine isolates nationally in 2018: four from New South Wales, three from Victoria, and two from Queensland.

The proportion of isolates resistant to penicillin in non-remote Australia ranged from 8.8% in non-remote Northern Territory to 44.1% in South Australia. In remote Northern Territory penicillin resistance rates remain low (1.9%), and higher in remote Western Australia (6.5%).

The proportion of isolates resistant to ciprofloxacin in non-remote Australia ranged from 10.3% in non-remote Northern Territory to 48.3% in South Australia. Ciprofloxacin resistance rates remain comparatively low in remote Northern Territory (1.9%) and remote Western Australia (4.6%).

Keywords: antimicrobial resistance, disease surveillance, gonococcal infection, *Neisseria gonorrhoeae*

Introduction

Antimicrobial resistance (AMR) in *Neisseria gonorrhoeae* (NG) is a threat to global health security, with the emergence and spread of multidrug-resistant gonorrhoea predicted to pose significant collateral health and financial costs.¹ In Australia, increasing gonococcal infection

rates,² the emergence of azithromycin resistance, and new reports of novel ceftriaxone-resistant strains with international spread,³ are key concerns and remain the focus of the National Neisseria Network (NNN). The NNN is a collaborative network of jurisdictional *Neisseria* reference laboratories that performs phenotypic and genotypic testing of clinical isolates of

pathogenic *Neisseria*. Clinical isolates referred to NNN laboratories from public and private sector laboratories represent as wide a section of the community as possible. The Australian Gonococcal Surveillance Programme (AGSP) is a key activity of the NNN which has continuously monitored NG AMR susceptibility since 1981; it is the longest, continually running, national surveillance system for gonococcal AMR.

In Australia, gonorrhoea notifications increased by 80% (from 65.5 to 118.0 per 100,000) in the five years 2013 to 2017. Increases were greater in males (91%) than in females (56%), with notification rates in 2016 remaining higher in males (174.2 per 100,000) than in females (61.8 per 100,000).²

In the five years 2013 to 2017, gonorrhoea annual notification rates in the Aboriginal and Torres Strait Islander (ATSI) population decreased by 19%; the 2017 gonorrhoea notification rate in the ATSI population remains markedly higher (6.6 times) than in the non-Indigenous population (627.5 per 100,000 versus 95.6 per 100,000) and is highest in remote and very remote areas (1,444 per 100,000; i.e., 30 times greater than the non-Indigenous population).² In contrast to non-remote Australia, NG AMR in remote regions remains low in locally acquired infections, with the recommended therapeutic strategy based on oral penicillin.⁴

Ceftriaxone and azithromycin dual therapy is recommended in Australia,⁵ having been introduced in 2014 in an attempt to forestall resistance to ceftriaxone; this was followed by both a steady decline in the proportion of isolates with raised MIC values to ceftriaxone, but also an increase in the proportion of isolates resistant to azithromycin, following rapid emergence of azithromycin resistance in South Australia in early 2016.⁶

Paradoxically, the current heightened global awareness of AMR, and increasing disease notification rates reported in Australia and elsewhere,^{2,7–10} have coincided with increased

use of nucleic acid amplification testing (NAAT) for diagnosis, replacing bacterial culture and antimicrobial susceptibility testing (AST). In remote regions in Australia, NAAT is used to detect penicillin resistance^{11,12} in NAAT positive samples for NG; this is the first documented use of routine molecular testing for NG AMR detection and surveillance, and these data continue to inform local treatment guidelines.¹²

The World Health Organization (WHO) 2018 Report on Global STI Surveillance estimates 87 million new NG infections annually worldwide in those aged 15–49 years, with the highest burden occurring in the Asia Pacific regions.¹³ Additional to this high disease burden in the Asia Pacific, the WHO Gonococcal Antimicrobial Surveillance Programme data indicate high levels of gonococcal AMR with significant gaps in surveillance. Furthermore, unregulated antimicrobial use in these regions provides ideal conditions for the development of AMR.¹⁴ The emergence of NG AMR in Australia has long been influenced by the introduction of multi-resistant strains from overseas.¹⁵ The importation and spread of resistant gonococcal strains, and/or new resistance developing, remains an ongoing concern.

Strategies for treating and controlling gonorrhoea are based on regimens effecting cure in a minimum of 95% of cases. Surveillance data of antibiotics in clinical use are critical to monitor AMR, to detect imported or novel resistance, and to inform treatment guidelines.¹⁶ The WHO has called for enhanced surveillance as a fundamental component of its Global Action Plan to control the spread and impact of gonococcal AMR.¹⁷

Methods

The NNN AMR data for gonococcal isolates are collated for the AGSP quarterly and annual reports. All confirmed cases of gonorrhoea in Australia are notifiable to the National Notifiable Diseases Surveillance System (NNDSS). The number of isolates tested by the NNN and reported by the AGSP represents a proportion

of the total number of cases reported to the NNDSS. The NNN laboratories test gonococcal isolates for susceptibility to penicillin; ceftriaxone; ciprofloxacin; azithromycin; spectinomycin and tetracycline, using previously-described standardised methodology to determine the MIC values.¹⁸ The MIC value is the lowest antibiotic concentration that inhibits *in vitro* growth under defined conditions. The AGSP conducts a program-specific quality assurance program.¹⁹

Gonococcal AST data from each jurisdiction are submitted quarterly to the coordinating laboratory (the Neisseria Reference Laboratory and WHO Collaborating Centre for Sexually Transmitted Infections and Antimicrobial Resistance, Sydney) which collates the data for reporting. Where available, the AGSP collects data on the sex of the patient, country of acquisition, and site of isolation of gonococcal isolates. Data collected across jurisdictions are predominantly from urban centres. Data from the Northern Territory and Western Australia are further divided into non-remote and remote regions determined at the jurisdictional level and coincident with antibiograms and consequent therapeutic recommendations.

Results

Number of gonococcal isolates and infections

NNN laboratories tested 9,006 NG isolates in 2018, representing the highest annual total of isolates ever assessed by the AGSP. There were 30,858 gonococcal infections notified to the NNDSS in 2018, similarly representing the highest number of annual gonococcal disease notifications since records commenced in 1991 (Figure 1).²⁰ Isolates from 29% of all cases of gonorrhoea notified to the NNDSS were tested by the NNN laboratories (Table 1); this rate equals the proportion tested in 2015–2017 but is lower than the rate from 2008–2014 (31% – 42%). The reduced referral rate reflects decreasing reliance on culture in Australia for NG diagnosis given the widespread uptake of NAAT.

Referred Isolates

There were 7,415 isolates from males (82.3%) and 1,547 (17.2%) from females (Table 2). 44 isolates were from patients where gender was not recorded. The proportion of gonococcal isolates from males and females has remained stable over recent years (2009–2017), ranging between 17 and 20% for women and 80 and 83% for men. The infected site was reported as ‘other’ or not specified for 83 isolates from males and 42 isolates from females (Table 2). Isolates from urine samples were regarded as genital tract isolates.

Antibiotic susceptibility patterns

As in previous years, gonococcal AMR patterns differed by state and territory (Table 3).

Ceftriaxone

Gonococcal isolates with decreased susceptibility to ceftriaxone (MIC values ≥ 0.06 mg/L) have been detected in Australia since 2001; the proportion increased to 4.4% in 2012, before doubling to 8.8% in 2013. From 2014, coincident with the introduction of dual ceftriaxone and azithromycin therapy, the proportion of isolates with decreased susceptibility to ceftriaxone fell annually to 1.06% in 2017 before increasing slightly to 1.73% in 2018 (see Table 4 and Table 5). From 2010, the proportion of isolates with an MIC value of ≥ 0.125 mg/L increased from 0.1% to 0.6% in 2013–2014 and then decreased to 0.04% in 2017. In 2018, an increase to 0.06% was observed as shown in Table 5. Gonococcal isolates from two patients from Queensland, not determined to be contacts, were found to be extensively drug-resistant (XDR) with ceftriaxone MIC values of 0.50 mg/L, high-level resistance to azithromycin (MIC ≥ 256 mg/L) and resistance to penicillin and ciprofloxacin. One had a history of travel to South East Asia.

Azithromycin

Nationally, in 2018, 6.2% of isolates exhibited azithromycin resistance (MIC ≥ 1.0 mg/L) (Table 3), decreasing from 9.3% reported in 2017.

Figure 1: Number of gonococcal disease cases reported to the National Notifiable Diseases Surveillance System compared with *N. gonorrhoeae* isolates available for laboratory testing by the Australian Gonococcal Surveillance Programme, Australia, 1991–2018

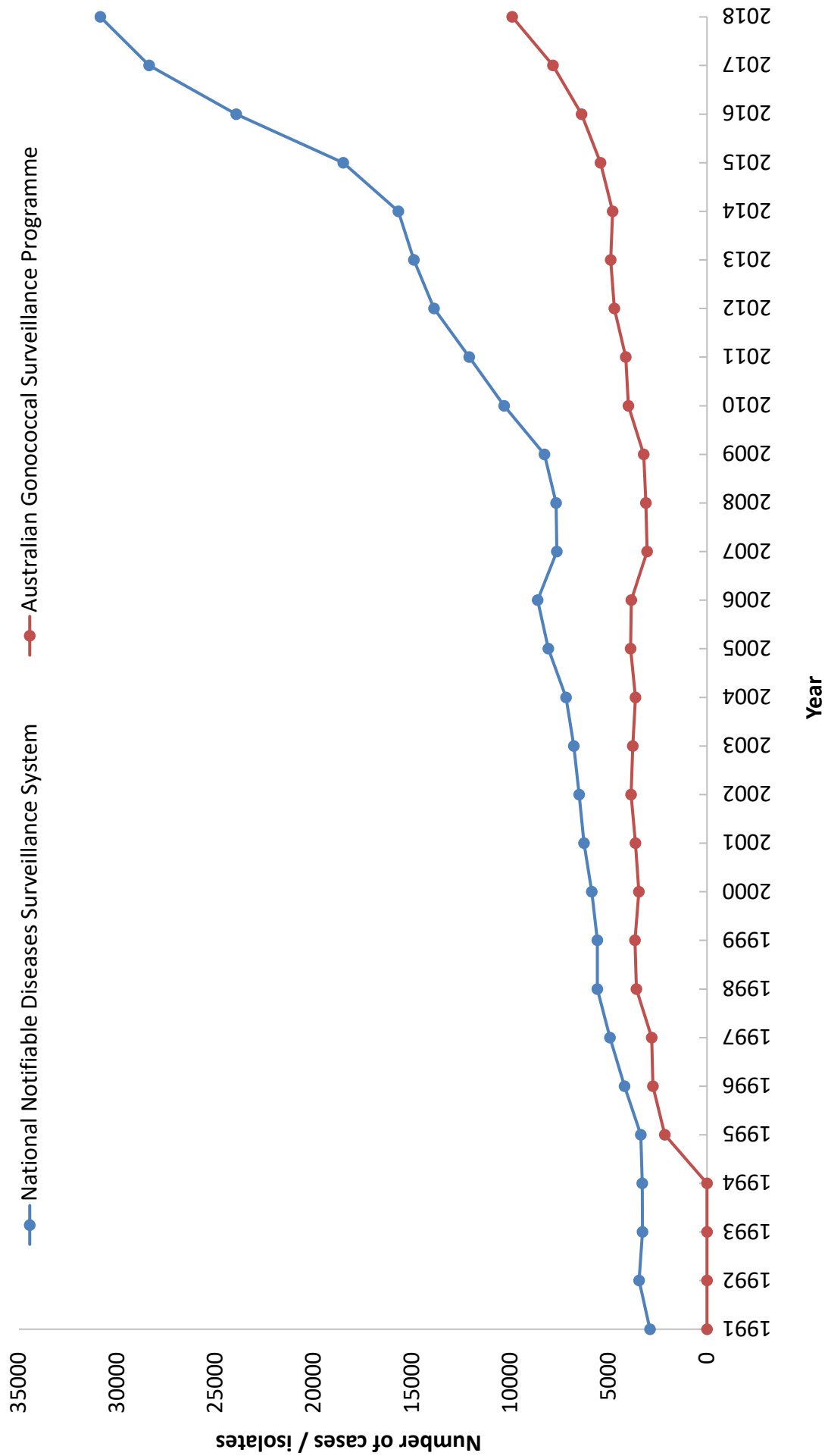


Table 1: Number of Australian Gonococcal Surveillance Programme gonococcal isolates tested as a proportion of National Notifiable Diseases Surveillance System gonorrhoea notifications, Australia, 2018, by state or territory

State or territory	Number of isolates tested	Number of cases notified	Isolates Tested/Notifications (%)
Australian Capital Territory	206	330	62
New South Wales	3,535	10,559	33
Northern Territory	225	2,124	11
Queensland	1,375	4,906	28
South Australia	231	1,288	18
Tasmania	55	149	37
Victoria	2,619	8,091	32
Western Australia	760	3,411	22
Australia	9,006	30,858	29

Table 2: Gonococcal isolates, Australia, 2018, by sex, site and jurisdiction tested.

Sex	Site	ACT	NSW	NT	Qld	SA	Vic	Tas	WA	AUSTRALIA
Male	Genital	65	1,614	167	628	106	1,111	20	405	4,116
	Rectal	51	854	2	270	50	659	11	99	1,996
	Pharynx	71	500	0	131	10	426	10	58	1,206
	DGI ^a	0	4	0	6	0	2	0	2	14
	Other/NS ^b	0	29	3	10	10	14	8	9	83
	Total	187	3,001	172	1,045	176	2,212	49	573	7,415
Female	Genital	11	378	52	302	43	299	3	179	1,267
	Rectal	0	14	0	7	3	7	0	1	32
	Pharynx	8	105	0	10	1	72	2	2	200
	DGI ^a	0	2	0	2	0	0	0	2	6
	Other/NS ^b	0	16	1	8	8	5	1	3	42
	Total	19	515	53	329	55	383	6	187	1,547
Unknown	Total	0	19	0	1	0	24	0	0	44
Total	Total	206	3,535	225	1,375	231	2,619	55	760	9,006

a DGI: Disseminated Gonococcal Infection

b NS: not specified

Table 3: Number and proportion (%) of gonococcal isolates with resistance to azithromycin, penicillin and ciprofloxacin and decreased susceptibility to ceftriaxone reported, Australia, 2018, by state or territory

Jurisdiction	Number of isolates tested 2018	Decreased susceptibility		Resistance					
		Ceftriaxone		Azithromycin		Penicillin		Ciprofloxacin	
		n	%	n	%	n	%	n	%
Australian Capital Territory	206	4	1.9	18	8.7	24	11.7	26	12.6
New South Wales	3,535	30	0.8	230	6.5	823	23.3	1,023	28.9
Queensland	1,375	18	1.3	68	4.9	284	20.7	379	27.6
South Australia ^a	231	3	1.3	7	3.0	52	44.1	57	48.3
Tasmania	55	4	7.3	3	5.5	14	25.5	20	36.4
Victoria	2,619	83	3.2	217	8.3	529	20.2	610	23.3
Northern Territory non-remote	68	0	0	1	1.5	6	8.8	7	10.3
Northern Territory remote	157	0	0	0	0	3	1.9	3	1.9
Western Australia non-remote	652	14	2.1	16	2.5	155	23.8	173	26.5
Western Australia remote	108	0	0	1	0.9	7	6.5	5	4.6
Australia	9,006	156	1.73	561	6.2	1,897	21.1	2,303	25.6

^a Number of isolates resistant to both penicillin and ciprofloxacin = 120

Table 4: Number and proportion (%) of gonococcal isolates with decreased susceptibility to ceftriaxone (MIC ≥ 0.06 mg/L), Australia, 2010 to 2018, by state or territory. Remote Western Australian data was de-aggregated from 2014.

Jurisdiction	Decreased susceptibility to ceftriaxone																	
	2010		2011		2012		2013		2014		2015		2016		2017		2018	
	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%	n	%
ACT	3	6.7	2	3.1	2	3.6	0	0	2	2.7	0	0.0	1	0.9	0	0	4	1.9
NSW	74	5.6	58	4.4	76	4.5	183	11.8	119	7.1	52	2.7	45	2.0	13	0.5	30	0.8
Qld	17	3.2	18	2.3	17	2.4	33	4.9	21	3.2	7	1.0	32	3.7	11	0.9	18	1.3
SA	12	11.6	1	0.7	1	0.7	4	1.9	2	1.0	9	3.6	2	0.6	2	0.6	3	1.3
Tas	0	0	0	0	0	0	11	24.4	0	0	0	0	1	3.6	0	0	4	7.3
Vic	52	5.7	50	5.3	105	8.4	181	11.8	95	6.6	25	1.5	19	1.1	48	2.1	83	3.2
NT non-Remote	1	0.2	2	0.4	0	0	2	1.9	3	3.0	0	0	0	0	0	0	0	0
NT Remote	0	0	0	0	0	0	2	0.8	1	0.8	0	0	0	0	0	0	0	0
WA	17	5.2	3	0.7	6	1.2	13	2.7										
WA Urban/Rural									14	3.6	5	1.3	9	1.3	9	1.4	14	2.1
WA Remote									1	0.9	0	0	0	0	0	0	0	0
Australia	191	4.8	134	3.2	207	4.4	429	8.8	258	5.4	98	1.8	109	1.7	83	1.1	156	1.7

Table 5: Proportion (%) of gonococcal isolates tested in Australia with ceftriaxone MIC values at 0.06 mg/L and ≥ 0.125 mg/L, 2010–2018.

Ceftriaxone MIC mg/L	2010	2011	2012	2013	2014	2015	2016	2017	2018
0.06	4.80%	3.20%	4.10%	8.20%	4.80%	1.70%	1.65%	1.02%	1.67%
≥ 0.125	0.10%	0.10%	0.30%	0.60%	0.60%	0.10%	0.05%	0.04%	0.06%

Since 2012 the rate of azithromycin resistance in Australian NG isolates increased almost five-fold, as shown in Table 6. Rates of azithromycin-resistant NG were highest in the Australian Capital Territory (8.7%), Victoria (8.3%) and New South Wales (6.5%) as shown in Tables 3 and 6. In 2018, 9 isolates exhibited high level resistance to azithromycin (MIC value \geq 256 mg/L), four from New South Wales, three from Victoria and two from Queensland. As noted above, the two isolates from Queensland were extensively drug-resistant (XDR) strains with high ceftriaxone MIC values. An additional 21 isolates were detected with resistance to azithromycin, penicillin and ciprofloxacin. 4.1% of azithromycin-resistant isolates demonstrated penicillin and ciprofloxacin resistance.

Penicillin

Resistance to the penicillin group of antibiotics (penicillin, ampicillin and amoxicillin with or without clavulanic acid) in NG results from β -lactamase production (i.e., penicillinase) and/or the aggregation of chromosomally-controlled resistance mechanisms. These are denoted respectively as penicillinase-producing *N. gonorrhoeae* (PPNG) and chromosomally mediated resistance to penicillin (CMRP). Chromosomal resistance is defined as a penicillin MIC value \geq 1 mg/L.

In 2018, in Australia, 1,897 (21.1%) isolates were penicillin resistant, a proportional decrease from 2016 (32.5%), and 2017 (26.1%). The proportion of penicillin-resistant isolates fluctuated in the range 22.5 to 44% between 2008 and 2017. In 2018, a total of 1,004 (11.3%) isolates had CMRP and 893 (10.0%) were PPNG; 47.1% of penicillin-resistant isolates were PPNG.

Penicillin resistance in remote Australia

In 2018, there were 225 isolates tested from the Northern Territory with 157 derived from remote areas of NT (including Alice Springs, Katherine, Tennant Creek, and Arnhem Land region) and 68 from Darwin and surrounding urban areas (non-remote). In 2018, a total of

760 isolates were tested from Western Australia, with 108 obtained from remote regions and 652 from urban and suburban Perth (non-remote).

Of the 157 isolates from remote NT, only 3 (1.9%) were penicillin resistant (1 was PPNG). 6 isolates (8.8%) from Darwin and surrounding urban areas were penicillin resistant (4 were PPNG) (Table 3). Of the 108 isolates from remote Western Australia, 7 (6.5%) were penicillin resistant, with 6 being PPNG. No isolate from remote NT or WA demonstrated decreased susceptibility to ceftriaxone.

Quinolones

The AGSP uses ciprofloxacin as the representative quinolone. Ciprofloxacin resistance is defined as MIC \geq 1 mg/L. In 2018, there were 2,303 ciprofloxacin resistant isolates (25.6%) (Table 3). Since 2008, when 54% of isolates tested ciprofloxacin resistant, the resistance rate has progressively declined.

Tetracyclines

To facilitate accurate reporting of NG tetracycline resistance in Australia, from 2018 NNN reference laboratories have performed tetracycline MIC testing where possible. This replaces historical testing for high level tetracycline testing which was reported by the NNN as an epidemiological marker for plasmid mediated resistance since inception. Tetracycline resistance is defined as an MIC value \geq 2 mg/L and utilises various mechanisms including plasmid-mediated resistance. The previously employed methods only detected high-level plasmid-mediated tetracycline-resistant *N. gonorrhoeae* (TRNG) (MIC value \geq 16 mg/L). Whilst tetracyclines are not a recommended treatment for gonorrhoea and are rarely, if ever, used for treatment of gonorrhoea in Australia, there has been recent interest in the proportion of tetracycline resistance. Nationally, 2,493 isolates were tested and 32% (797/2,493) were tetracycline resistant. Tetracycline resistance data are presented by jurisdiction and aggregated for Australia as shown in Table 7.

Table 6: Number and proportion (%) of gonococcal isolates with resistance to azithromycin (MIC ≥ 1.0 mg/L), Australia, 2012 to 2018, by state or territory.

State or territory	Azithromycin resistance													
	2012		2013		2014		2015		2016		2017		2018	
	n	%	n	%	n	%	n	%	n	%	n	%	n	%
ACT	0	0	1	2.2	7	9.3	0	0	8	7.1	3	2.1	18	8.7
NSW	9	0.5	14	0.9	33	2.0	43	2.3	82	3.6	261	9.3	230	6.5
Qld	15	2.1	38	5.7	23	3.5	42	5.8	10	1.2	61	4.9	68	4.9
SA	1	0.7	6	2.8	1	0.5	7	2.8	68	19.5	46	12.8	7	3.0
Tas	0	0	0	0	1	3.3	1	4.3	4	14.3	5	9	3	6
Vic	34	2.7	35	2.3	33	2.3	30	1.8	93	5.4	304	13.5	217	8.3
NT Urban & Rural	0	0	1	1.0	0	0	0	0	1	1.9	1	1.7	1	1.5
NT Remote	0	0	0	0	0	0	0	0	0	0	1	0.6	0	0.0
WA Urban and Rural	3	0.6	9	1.9	21	5.3	15	3.8	51	7.6	40	6.4	16	2.5
WA Remote	0	0	0	0	0	0	0	0	1	0.8	4	3.4	1	0.9
Australia	62	1.3	104	2.1	119	2.5	138	2.6	318	5.0	726	9.3	561	6.2

Spectinomycin

In 2018, all isolates tested were susceptible to spectinomycin.

Discussion

The WHO recommends that treatment regimens for gonorrhoea are based on epidemiological surveillance of the distribution and extent of AMR, and that a resistance rate of 5% or more is the nominal threshold for change of treatment recommendations.¹⁶ The AGSP has continuously monitored gonococcal AMR in Australia since 1981, and has established and coordinated quality assurance, and quality control, for gonococcal AMR testing with the AGSP External Quality Assurance Program, and the development of the WHO NG reference strains, thus ensuring the quality of the AGSP data.^{19,21} In 2018 the NNN examined 9,006 clinical isolates for susceptibility testing to ceftriaxone, azithromycin, ciprofloxacin, penicillin and high-level resistance to tetracycline. These isolates were referred from both the public and private health sectors, constituting a comprehensive sample of about one-third of all notifications nationally. Coincident with the increases in NG disease rates in Australia,² there has been an increase in the numbers of gonococcal isolates tested for AMR. However, the proportion remains relatively unchanged at about 30%.

Both in Australia and internationally, the monitoring of ceftriaxone and azithromycin MIC values is the primary focus of surveillance for gonococcal AMR. With regard to ceftriaxone, MIC values ≥ 0.06 mg/L are reported to have decreased susceptibility. The AGSP proportion of isolates with decreased susceptibility to ceftriaxone has steadily and substantially declined since 2013 from 8.8% to 1.7% in 2018 (Table 4). However little reassurance should be taken from this, as multidrug-resistant strains with high-level resistance to ceftriaxone have been reported from Asia, Europe and Australia in recent years.^{22–25}

In 2017 the AGSP identified two new multidrug-resistant NG isolates in Australia. These strains were phenotypically similar to the ceftriaxone-resistant strain first reported in Japan in 2015 (FC428), and similar strains in Denmark (GK124) and Canada (47,707).³ Further investigations, in collaboration by the NNN with international colleagues, found that, on bioinformatic analysis, there was close genetic relatedness amongst other phenotypically similar isolates from Japan and Canada providing further evidence of international transmission of this ceftriaxone-resistant *N. gonorrhoeae* strain.³

In 2018 two isolates of global concern were detected by the AGSP and reported by the Australian Government to the World Health Organization due to their international significance. These two isolates, with ceftriaxone MIC values of 0.50 mg/L, high-level resistance to azithromycin (MIC value ≥ 256 mg/L), and resistance to penicillin and ciprofloxacin were isolated from patients, not determined to be contacts, from Queensland. One had a history of travel to South East Asia. At the same time an isolate with a similar phenotype was identified in England, also with a history of travel to South East Asia.²⁶ Again, the NNN, through international collaboration, this time with UK colleagues through the WHO Collaborating Centre Network, worked to investigate the genotypic analysis of these three isolates and determined that they represent a single gonococcal clone, the A2543 clone.²⁷ In all likelihood this A2543 clone is circulating, but undetected due to gaps in NG AMR surveillance internationally.²⁷

Remote populations of Australia, which are predominantly Aboriginal and Torres Strait Islander, have low rates of AMR despite very high rates of disease, but require continued vigilance with monitoring of AMR in NG using molecular and culture-based surveillance strategies.

In 2013, high-level resistance (HLR; MIC value ≥ 256 mg/L) to azithromycin in gonococci was reported for the first time in Australia in four strains, two with suspected contact in China.²⁸ Since then there have been only sporadic reports

Table 7: Number and proportion (%) of gonococcal isolates with resistance to tetracycline (MIC ≥ 2 mg/L), Australia, 2018, by state or territory

State or Territory	Number of isolates tested	Resistance MIC ≥ 2 mg/L	
	2018	Tetracycline	
		n	%
Australian Capital Territory	190	16	8.4
New South Wales	88	59	67.0
Queensland	3	2	66.7
South Australia	2	1	50
Tasmania	53	9	17.0
Victoria	1,320	519	39.3
Northern Territory (non-remote)	17	3	17.6
Northern Territory (remote)	65	5	7.7
Western Australia (non-remote)	648	173	26.7
Western Australia (remote)	107	10	9.3
AUSTRALIA	2,493	797	32.0

of HLR to azithromycin; however, there were nine such strains in 2018. Additionally, there were 23 isolates (including the XDR strains described above) that were resistant to azithromycin, penicillin and ciprofloxacin. The proportion of azithromycin resistant isolates that were also resistant to penicillin and ciprofloxacin was 4.1%. Continued close observation is ongoing as evidence of coevolving cephalosporin and azithromycin resistance is being observed outside Australia and is of significant concern.²⁹

Another important and concerning finding by the AGSP in 2018 is the increase in isolates with low level resistance to azithromycin in all jurisdictions of Australia, excepting the ACT and Northern Territory (Table 6). In remote WA there were four azithromycin resistant strains reported. Until recently azithromycin resistance in Australia in NG has remained relatively low at 1.3–2.6% over the years 2012–2015 but has then increased from 5% in 2016 to 9.3% in 2017

(Table 6). In South Australia in 2016, azithromycin resistance in NG significantly increased ($p < 0.0001$) from less than 5% in the latter half of 2015 to 26% in the first half of 2016.³⁰ Overall in 2016, there were 68/349 (19.5%) strains in South Australia that were azithromycin resistant with MIC values in the range 1.0 mg/L to 8.0 mg/L. Enhanced surveillance was conducted, and one treatment failure was reported in a patient treated with azithromycin single agent therapy.⁶ A review and change of the South Australian gonococcal treatment guidelines followed.⁶ In 2018 azithromycin resistance was highest in the Australian Capital Territory (8.7%), Victoria (8.3%) and New South Wales 6.5%). Globally there have been increasing reports of azithromycin resistance.³¹

The recent reports of international spread of NG with resistance to ceftriaxone,²⁶ and the emergence of azithromycin resistance, heighten concerns about the future treatment strategies

for NG AMR. Public health strategies promoting primary prevention of gonorrhoea and other sexually transmissible infections are urgently required, and NG vaccine development is a research priority to control this disease. This report underscores the importance of bacterial culture and antimicrobial susceptibility testing of NG for clinical management, detection of resistance and novel resistant strains, AMR surveillance, and test of cure. Clinicians should note and consider travel history given the association with NG AMR.

The WHO Global Action Plan states that disease control strategies and the understanding of the global scope of AMR need to continue to be informed by surveillance programs of AMR, nationally and internationally.¹⁷ The ongoing need for close and enhanced monitoring of gonococcal AMR can be supported, but not replaced, by molecular based assays and strain specific assays can be used for routine and sentinel site surveillance in high risk populations. The data are critically important to inform therapeutic strategies, to monitor for the presence and spread of resistance and to detect instances of treatment failure.

Acknowledgements

The NNN is supported by the Commonwealth Department of Health to provide the AGSP. We thank the many laboratories, private and public, throughout Australia for referral of isolates for testing, and Dr David Speers for the communication regarding the PPNG assay data from WA.

Members of the NNN in 2018 (and to whom isolates should be referred) were: John Bates and Vicki Hicks (Queensland Public Health Microbiology, Forensic and Scientific Services, Coopers Plains, Queensland); Athena Limnios, Tiffany Hogan, Ratan Kundu, Rodney Enriquez, Jasmin El-Nasser and Monica M. Lahra (New South Wales Health Pathology Department of Microbiology, The Prince of Wales Hospital, Randwick, New South Wales and School of Medical Sciences, Faculty of Medicine, the University of New South Wales, Kensington,

NSW Australia); Kerrie Stevens, Samantha Tawil, and Benjamin P. Howden (The Microbiological Diagnostic Unit (PHL), Department of Microbiology and Immunology, Peter Doherty Institute for Infection and Immunity, The University of Melbourne, Parkville, Victoria); Andrew Lawrence and Judith Holds (SA Pathology); Julie Pearson; Jane Bew and David Speers (Department of Microbiology and Infectious Diseases, PathWest Laboratory Medicine, Fiona Stanley Hospital, Western Australia); Belinda McEwan (Department of Microbiology and Infectious Diseases, Royal Hobart Hospital, Hobart, Tasmania); Kevin Freeman and Microbiology Staff (Microbiology Laboratory, Territory Pathology, Royal Darwin Hospital, Tiwi, Northern Territory); and Susan Bradbury and Peter Collignon (Microbiology Department, The Canberra Hospital, Garran, Australian Capital Territory).

Author details

Monica M Lahra^{1,2,3}

Rodney Enriquez^{1,2}

C. R. Robert George^{1,4}

1. The National Neisseria Network, Australia
2. Neisseria Reference Laboratory and World Health Organisation Collaborating Centre for STD, Sydney. New South Wales Health Pathology, Microbiology, The Prince of Wales Hospital, Randwick, NSW Australia.
3. School of Medical Sciences, Faculty of Medicine, The University of New South Wales, 2053 Australia
4. New South Wales Health Pathology, Microbiology, John Hunter Hospital, New Lambton Heights, NSW Australia.

Corresponding author:

Professor Monica M Lahra, World Health Organization Collaborating Centre for STD,

Sydney and, Neisseria Reference Laboratory, Microbiology Department, SEALS, The Prince of Wales Hospital, Randwick, NSW, 2031. School of Medical Sciences, Faculty of Medicine, the University of New South Wales, NSW 2050 Australia.

Telephone: +61 2 9382 9050.

Facsimile: +61 2 9382 9210.

Email: monica.lahra@health.nsw.gov.au

References

1. Centers for Disease Control and Prevention. *Antibiotic Resistance Threats in the United States*. Atlanta: US Department of Health and Human Services, Centers for Disease Control and Prevention, 2013.
2. Kirby Institute. *HIV, viral hepatitis and sexually transmissible infections in Australia: Annual Surveillance Report 2018*. Sydney: Kirby Institute, UNSW Sydney, 2018. Available from: https://kirby.unsw.edu.au/sites/default/files/kirby/report/KI_Annual-Surveillance-Report-2018.pdf.
3. Lahra MM, Martin I, Demczuk W, Jennison AV, Lee KI, Nakayama SI et al. Cooperative recognition of internationally disseminated ceftriaxone-resistant *Neisseria gonorrhoeae* strain. *Emerg Infect Dis*. 2018;24(4). <https://doi.org/10.3201/eid2404.171873>.
4. Lahra MM, Enriquez R. Australian Gonococcal Surveillance Programme Annual Report, 2016. *Commun Dis Intell (2018)*. 2018;42. pii: S2209-6051(18)00013-1.
5. Australasian Sexual Health Alliance. Australian STI management guidelines for use in primary care: Gonorrhoea. [Internet.] Australasian Sexual Health Alliance, 2018. Available from: <http://www.sti.guidelines.org.au/sexually-transmissible-infections/gonorrhoea>.
6. Lahra MM, Ward A, Trembizki E, Her-manson J, Clements E, Lawrence A et al. Treatment guidelines after an outbreak of azithromycin-resistant *Neisseria gonorrhoeae* in South Australia. *Lancet Infect Dis*. 2017;17(2):133–4.
7. Family Planning Association. Sexually transmitted infections factsheet. [Internet.] United Kingdom: Family Planning Association, 2016. Available from: <https://www.fpa.org.uk/factsheets/sexually-transmitted-infections>.
8. Centers for Disease Control and Prevention. *2016 Sexually Transmitted Diseases Surveillance Report*. Atlanta: U.S. Department of Health and Human Services, Centers for Disease Control and Prevention, 2017.
9. European Centre for Disease Prevention and Control. Annual Epidemiological Report 2016 – Gonorrhoea. [Internet.] Stockholm: European Centre for Disease Prevention and Control, 2016. Available from: https://ecdc.europa.eu/sites/portal/files/documents/Gonorrhoea%20AER_0.pdf.
10. Government of Canada. Gonorrhoea. [Internet.] Ottawa: Government of Canada, 2017. Available from: <https://www.canada.ca/en/public-health/services/diseases/gonorrhoea.html>.
11. Goire N, Freeman K, Tapsall JW, Lambert SB, Nissen MD, Sloots TP et al. Enhancing gonococcal antimicrobial resistance surveillance: a real-time PCR assay for detection of penicillinase-producing *Neisseria gonorrhoeae* by use of noncultured clinical samples. *J Clin Microbiol*. 2011;49(2):513–8.
12. Speers DJ, Fisk RE, Goire N, Mak DB. Non-culture *Neisseria gonorrhoeae* molecular penicillinase production surveillance demonstrates the long-term success of empirical dual therapy and informs gonorrhoea management guidelines in a highly endemic setting. *J Antimicrob Chemother*.

- 2014;69(5):1243–7.
13. World Health Organization (WHO). *Report on global sexually transmitted infection surveillance 2018*. Geneva: WHO, 2018.
 14. Lahra MM, Lo YR, Whiley DM. Gonococcal antimicrobial resistance in the Western Pacific Region. *Sex Transm Infect*. 2013;89(Suppl 4):iv19–23.
 15. Tapsall JW, Limnios EA, Murphy D. An analysis of trends in antimicrobial resistance in *Neisseria gonorrhoeae* isolated in Australia, 1997 – 2006. *J Antimicrob Chemother*. 2008;61(1):150–5.
 16. Tapsall J, World Health Organization (WHO) Anti-Infective Drug Resistance Surveillance and Containment Team. *Antimicrobial resistance in Neisseria gonorrhoeae*. Switzerland: World Health Organization, 2001. Available from: <https://apps.who.int/iris/handle/10665/66963>.
 17. World Health Organization (WHO), Department of Reproductive Health and Research. *Global action plan to control the spread and impact of antimicrobial resistance in Neisseria gonorrhoeae*. Switzerland: WHO, 2012.
 18. Bell SM, Pham JN, Rafferty DL, Allerton JK. *Antibiotic susceptibility testing by the CDS method: A manual for medical and veterinary laboratories*. 8th ed. Kogarah, NSW: South Eastern Area Laboratory Services, 2016.
 19. Tapsall JW. Use of a quality assurance scheme in a long-term multicentric study of antibiotic susceptibility of *Neisseria gonorrhoeae*. *Genitourin Med*. 1990;66(1):8–13.
 20. Australian Government Department of Health. National Notifiable Diseases Surveillance System. [Internet.] Canberra: Australian Government, Department of Health, 2018. [Accessed 16 Apr 2018.] Available from: [http://www9.health.gov.au/cda/](http://www9.health.gov.au/cda/source/cda-index.cfm)
 21. Unemo M, Fasth O, Fredlund H, Limnios A, Tapsall J. Phenotypic and genetic characterization of the 2008 WHO *Neisseria gonorrhoeae* reference strain panel intended for global quality assurance and quality control of gonococcal antimicrobial resistance (AMR) surveillance for public health purposes. *J Antimicrob Chemother*. 2009;63(6):1142–51.
 22. Ohnishi M, Golparian D, Shimuta K, Saika T, Hoshina S, Iwasaku K et al. Is *Neisseria gonorrhoeae* initiating a future era of untreatable gonorrhoea?: Detailed characterization of the first strain with high-level resistance to ceftriaxone. *Antimicrob Agents Chemother*. 2011;55(7):3538–45.
 23. Unemo M, Golparian D, Nicholas R, Ohnishi M, Gallay A, Sednaoui P. High-level cefixime- and ceftriaxone-resistant *Neisseria gonorrhoeae* in France: Novel penA mosaic allele in a successful international clone causes treatment failure. *Antimicrob Agents Chemother*. 2012;56(3):1273–80.
 24. Cámara J, Serra J, Ayats J, Bastida T, Carnicer-Pont D, Andreu A et al. Molecular characterization of two high-level ceftriaxone-resistant *Neisseria gonorrhoeae* isolates detected in Catalonia, Spain. *J Antimicrob Chemother*. 2012;67:1858–60.
 25. Lahra MM, Ryder N, Whiley DM. A new multidrug-resistant strain of *Neisseria gonorrhoeae* in Australia. *N Engl J Med*. 2014;371(19):1850–1.
 26. European Centre for Disease Prevention and Control. Rapid Risk Assessment: Extensively drug-resistant (XDR) *Neisseria gonorrhoeae* in the United Kingdom and Australia – 7 May 2018. [Internet.] Stockholm: ECDC, 2018.
 27. Jennison AV, Whiley D, Lahra MM, Graham RM, Cole MJ, Hughes G et al. Genetic relatedness of ceftriaxone-resistant and high-level

azithromycin resistant *Neisseria gonorrhoeae* cases, United Kingdom and Australia, February to April 2018. *Euro Surveill.* 2019;24(8):
[https://doi.org/10.2807/1560-7917.](https://doi.org/10.2807/1560-7917.ES.2019.24.8.1900118)
ES.2019.24.8.1900118.

28. Stevens K, Zaia A, Tawil S, Bates J, Hicks V, Whiley D et al. *Neisseria gonorrhoeae* isolates with high-level resistance to azithromycin in Australia. *J Antimicrob Chemother.* 2015;70(4):1267–8.
29. Whiley DM, Lahra MM, Unemo M. Prospects of untreatable gonorrhoea and ways forward. *Future Microbiol.* 2015;10(3):313–6.
30. Lahra MM, Enriquez RP. Australian Gonococcal Surveillance Programme, 1 July to 30 September 2016. *Commun Dis Intell Q Rep.* 2017;41(1):E109–10.
31. Unemo M. Current and future antimicrobial treatment of gonorrhoea – the rapidly evolving *Neisseria gonorrhoeae* continues to challenge. *BMC Infect Dis.* 2015;15:364.